

Identification of a Mo–Fe–S Cluster on NifEN by Mo K-Edge Extended X-ray Absorption Fine Structure

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The iron–molybdenum cofactor (FeMo-co) is the complex metal–sulfur cofactor found within the active site of the molybdenum–iron protein (MoFe protein) component of the nitrogenase enzyme complex.¹ FeMo-co is among the most complex metal–sulfur cofactors known in biology and is composed of 7 Fe, 9 S, Mo, X (C, N, or O) and *R*-homocitrate.² This cofactor is required for the nitrogenase MoFe protein to catalyze the reduction of N₂ to NH₃ in nitrogen-fixing organisms, such as *Azotobacter vinelandii* or *Rhodospirillum rubrum*, and thus is essential for the input of nitrogen into the biosphere.

Compared to simpler clusters, the large size and high complexity of FeMo-co is supported by a biosynthetic pathway that is more complex than that required for building simpler clusters such as [Fe₂S₂], [Fe₃S₄], or [Fe₄S₄]. A series of combined genetic and biochemical studies have shown that multiple proteins, mostly encoded by nitrogen fixation (*nif*) genes, are involved in the biosynthesis of FeMo-co.³ FeMo-co is assembled outside of the MoFe protein and then is inserted into the apo-MoFe protein to constitute an active nitrogenase capable of nitrogen reduction. A full description of the biosynthetic pathway and the relevant proteins is available in recent reviews.³

NifB, NifEN, and the nitrogenase Fe protein (NifH) are minimally essential for the biosynthesis of FeMo-co, and novel enzymatic reactions for assembling and modifying Fe–S clusters are hypothesized for NifB and NifEN. NifB, a member of the “SAM-radical enzyme” family,⁴ has recently been isolated and used to start the assembly of FeMo-co in vitro using Fe²⁺ and S²⁻ as its initial substrates.⁵ NifB catalyzes the formation of NifB-co, an isolable low-molecular-weight intermediate of the FeMo-co biosynthesis pathway that is hypothesized to provide the majority of the Fe and S atoms of FeMo-co, but not Mo.⁶ In experiments using anoxic native electrophoretic analysis, the NifEN complex, which is homologous to the MoFe protein,⁷ has been shown to bind NifB-co.⁸ NifEN serves as a molecular scaffold where some of the steps for the assembly of FeMo-co occur. The NifEN protein is hypothesized to convert NifB-co to FeMo-co by the addition of (i) possibly more Fe and S, (ii) Mo, and (iii) *R*-homocitrate in a series of reactions, some of which are dependent on NifH.^{9,10} There is also experimental evidence to suggest that NifH may bind a FeMo-co precursor at some stage and that NifX may be involved in the attachment of *R*-homocitrate.⁹ NafY binds the completed FeMo-co with high affinity, and it is predicted to increase the efficiency of inserting it into the apo-MoFe protein.¹¹

As mentioned above, the combined action of NifEN and NifH have been shown to be required for conversion of NifB-co into

FeMo-co. The mechanism of this conversion that occurs within the NifEN protein is largely unknown. Among the key questions are (i) what are the structures of the Fe–S and Mo substrates? (ii) how many and what types of metal clusters are present on NifEN? (iii) how do the substrates interact with the pre-existing clusters? The NifEN complex as purified from a Δ *nifHDK* strain of *A. vinelandii* (DJ1041) by an improved procedure contains 24 Fe per tetramer,¹² but the distribution of these Fe atoms in the different clusters has not been well characterized. At minimum, NifEN has two [Fe₄S₄] clusters of unknown function and an Fe–S cluster that serves as a FeMo-co precursor.¹³ In addition, the as-purified NifEN has also been determined to contain substoichiometric amounts of bound Mo.¹² Biochemical studies have shown that the bound Mo is suitable for in vitro FeMo-co synthesis. In this study, the ligand environment of the Mo bound on the as purified NifEN is examined by Mo K-edge X-ray absorption spectroscopy (XAS).

The Mo K near-edge spectrum of NifEN is presented in Figure 1A. The quantity of Mo present, determined by integrating the Mo K α fluorescence at 20.5 keV incident energy, was 0.13 mM for 0.9 mM NifEN tetramer. The spectrum exhibits relatively little structure. When compared to the spectrum from *A. vinelandii* MoFe protein (Figure 1A,B), the two edges occur at similar energies, but the MoFe protein has additional edge structure around 20.020 keV while NifEN has a shoulder at 20.008 keV. This indicates that the Mo sites in NifEN and the MoFe protein have similar oxidation states but significantly different ligand fields.

Analysis of the extended X-ray absorption fine structure (EXAFS) data (Figure 1C,D) shows that the spectrum fits well to 3 Fe atoms at a distance of 2.68 Å and 3 S atoms at 2.35 Å. These distances are very similar to those observed for the nitrogenase M-center (FeMo-co),¹⁴ and they strongly suggest that the Mo is part of a cluster whose first two coordination spheres resemble a [MoFe₃S₄] cubane fragment as part (or whole) of some cluster on NifEN. Attempts to include non-sulfur Mo–O or Mo–N interactions did not give appreciably better fits. While the presence of short Mo–O ligands below 2.0 Å, consistent with a Mo=O double bond, could be excluded, it was not possible to ascertain the presence of longer Mo–O/N in the 2.0–2.3 Å range, as these correlate strongly with the Mo–S interaction over the *k*-range of the data.¹⁵

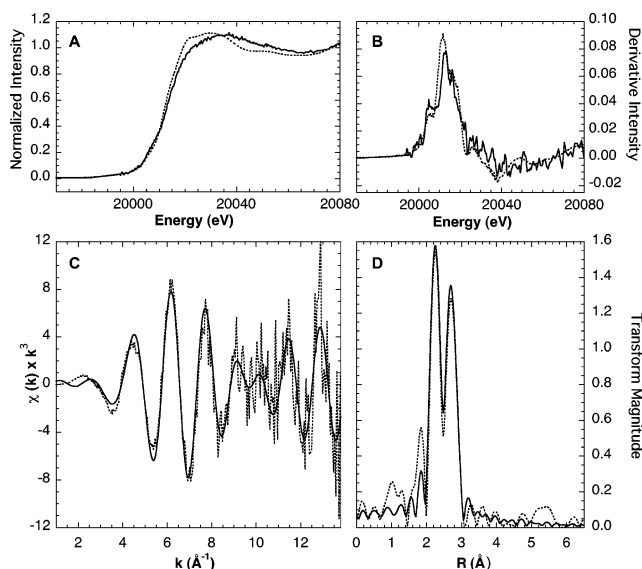
The presence of Mo in a Fe–S cluster environment raises the question of which cluster on NifEN contains the Mo. Possibilities for the Mo-containing cluster could involve (i) finished FeMo-co, (ii) a FeMo-co precursor containing Mo, or (iii) another separate cluster that may be permanent or transient. These three possibilities are discussed below.

First, the Mo bound to NifEN is unlikely to be part of finished FeMo-co, because the differences in the Mo K-edge spectrum of

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	N	R (Å)	σ^2 (Å ²)	ΔE_0 (eV)
Mo-Fe	3	2.684 (0.007)	0.0045 (0.0002)	-10.8 (2.0)
Mo-S	3	2.349 (0.002)	0.0030 (0.0003)	-6.3 (1.5)

Figure 1. Mo K-edge XANES and EXAFS recorded from of *A. vinelandii* NifEN. (A) XANES spectrum of NifEN (solid line) compared with *A. vinelandii* MoFe protein (broken line). (B) First derivative spectra from A. (C) k^3 weighted EXAFS spectrum of NifEN (broken line) and analysis (solid line). (D) Mo-Fe phase corrected Fourier transforms from C. (bottom panel) Fitting parameters, where N is the number of atoms, R the distance, σ^2 the Debye-Waller factor, and ΔE_0 the offset from the threshold energy. The fitting uncertainties are in brackets. In the presented fit, N has been constrained to a half-integer.

NifEN and the MoFe protein indicate that the ligand environment is not the same, possibly owing to a lack of homocitrate. Moreover, the NifEN protein was isolated from a strain lacking NifH, which is required for the final steps of FeMo-co biosynthesis.

Second, we consider it unlikely that the Mo is part of a NifB-co derivative bound on NifEN because of results from our laboratory on the transfer of a Fe-S cluster between NifEN and NifX. These experiments show that the NifX protein can extract an Fe-S cluster, designated as the VK-cluster, from NifEN charged with a NifB-co derivative.¹⁶ The NifX-bound VK-cluster can be used as a FeMo-co precursor in an in vitro FeMo-co synthesis reaction, indicating that the VK-cluster is the extracted form of the NifB-co derivative bound to NifEN. Metal quantification by inductively-coupled optical-emission spectroscopy showed that the VK-cluster contains virtually no Mo,¹⁶ which, in turn, suggests that the NifEN-bound NifB-co derivative does not contain Mo.¹⁷ We note that a better Mo EXAFS data set with longer k -range and lower noise levels should be able to demonstrate whether the Mo is part of finished FeMo-co or a NifB-co derivative through the presence or absence of the long-range 5 Å Mo-Fe interaction observed in MoFe protein.

If the Mo within NifEN is not part of a large cluster, it may be part of a simpler metal-sulfur cluster. The EXAFS data supports the existence of a $[\text{MoFe}_3\text{S}_{3+x}]$ cluster. The presence of an additional $[\text{MoFe}_3\text{S}_{3+x}]$ cluster on NifEN, separate from the $[\text{Fe}_4\text{S}_4]$ previously identified by resonance Raman measurements,^{13(a)} is not unreasonable to accommodate the reactions hypothesized to occur within NifEN to convert NifB-co into FeMo-co. An additional cluster would also be compatible with the number of Fe atoms determined for the as-purified NifEN complex.¹² A $[\text{MoFe}_3\text{S}_{3+x}]$ cluster on NifEN is consistent with the likely existence of a $[\text{Fe}_3\text{S}_4]$ cluster on NifEN. Previous EPR inspection of NifEN oxidized by either indigo carmine or thionine revealed a broad asymmetric isotropic signal at $g = 1.95$.¹³ EPR signals having similar line shape

are characteristic of $S = 1/2$ triferic $[\text{Fe}_3\text{S}_4]^{1+}$ clusters in proteins,¹⁸ and the reduced $[\text{Fe}_3\text{S}_4]^0$ state is known to be able to coordinate heterometals to complete a $[\text{MFe}_3\text{S}_4]$ cubane.¹⁹ Hence, a $[\text{Fe}_3\text{S}_4]$ center may serve as the initial Mo binding site either by reductive coupling to molybdate or from a reduced Mo species. The resulting $[\text{MoFe}_3\text{S}_4]$ within NifEN may well act as Mo donor for the bound NifB-co derived FeMo-co precursor in the protein complex. This hypothesis is currently being examined in our laboratory.

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Supporting Information Available: Experimental procedures and additional EXAFS fits. This material is available free of charge via the Internet at <http://pubs.acs.org>.

References

- (1) (a) Ludden, P. W.; Rangaraj, P.; Rubio, L. M. In *Catalysts for Nitrogen Fixation: Nitrogenases, Relevant Chemical Models, and Commercial Processes*; Smith, B. E., Richards, R. L., Newton, W. E., Eds.; Kluwer: Dordrecht, The Netherlands, 2004; pp 219–253. (b) Shah, V. K.; Brill, W. J. *Proc. Natl. Acad. Sci. U.S.A.* **1977**, *74*, 3249–3253.
- (2) (a) Einsle, O.; Tezcan, F. A.; Andrade, S. L.; Schmid, B.; Yoshida, M.; Howard, J. B.; Rees, D. C. *Science* **2002**, *297*, 1696–1700. (b) Hoover, T. R.; Robertson, A. D.; Cerny, R. L.; Hayes, R. N.; Imperial, J.; Shah, V. K.; Ludden, P. W. *Nature* **1987**, *329*, 855–857. (c) Chan, M. K.; Kim, J.; Rees, D. C. *Science* **1993**, *260*, 792–794.
- (3) (a) Dos Santos, P. C.; Dean, D. R.; Hu, Y.; Ribbe, M. W. *Chem. Rev.* **2004**, *104*, 1159–1174. (b) Rubio, L. M.; Ludden, P. W. *J. Bacteriol.* **2005**, *187*, 405–414.
- (4) Sofia, H. J.; Chen, G.; Hetzler, B. G.; Reyes-Spindola, J. F.; Miller, N. E. *Nucleic Acids Res.* **2001**, *29*, 1097–1106.
- (5) Curatti, L.; Ludden, P. W.; Rubio, L. M. *Proc. Natl. Acad. Sci. U.S.A.* **2006**, *103*, 5297–5301.
- (6) Shah, V. K.; Allen, J. R.; Spangler, N. J.; Ludden, P. W. *J. Biol. Chem.* **1994**, *269*, 1154–1158.
- (7) Brigle, K. E.; Weiss, M. C.; Newton, W. E.; Dean, D. R. *J. Bacteriol.* **1987**, *169*, 1547–1553.
- (8) Roll, J. T.; Shah, V. K.; Dean, D. R.; Roberts, G. P. *J. Biol. Chem.* **1995**, *270*, 4432–4437.
- (9) Rangaraj, P.; Ludden, P. W. *J. Biol. Chem.* **2002**, *277*, 40106–40111.
- (10) (a) Hu, Y.; Corbett, M. C.; Fay, A. W.; Webber, J. A.; Hodgson, K. O.; Hedman, B.; Ribbe, M. W. *Proc. Natl. Acad. Sci. U.S.A.* **2006**, *103*, 17125–17130. (b) Hu, Y.; Corbett, M. C.; Fay, A. W.; Webber, J. A.; Hodgson, K. O.; Hedman, B.; Ribbe, M. W. *Proc. Natl. Acad. Sci. U.S.A.* **2006**, *103*, 17119–17124.
- (11) Rubio, L. M.; Singer, S. W.; Ludden, P. W. *J. Biol. Chem.* **2004**, *279*, 19739–19746.
- (12) Soboh, B.; Igarashi, R. Y.; Hernandez, J. A.; Rubio, L. M. *J. Biol. Chem.* **2006**, *281*, 36701–36709.
- (13) (a) Goodwin, P. J.; Agar, J. N.; Roll, J. T.; Roberts, G. P.; Johnson, M. K.; Dean, D. R. *Biochemistry* **1998**, *37*, 10420–10428. (b) Goodwin, P. J. Ph.D. Dissertation, Virginia Polytechnic Institute, 1999. (c) Hu, Y.; Fay, A. W.; Ribbe, M. W. *Proc. Natl. Acad. Sci. U.S.A.* **2005**, *102*, 3236–3241.
- (14) Christiansen, J.; Tittsworth, R. C.; Hales, B. J.; Cramer, S. P. *J. Am. Chem. Soc.* **1995**, *117*, 10017–10024.
- (15) (a) George, G. N. *J. Biol. Inorg. Chem.* **1997**, *2*, 790–796. (b) Marginally improved fits could be obtained with combinations of Mo-O interactions at ~ 2.0 Å and ~ 2.3 Å, but these interactions correlate with each other and the Mo-S making unambiguous assignment impossible with the current data set (see Supporting Information).
- (16) Hernandez, J. A.; Igarashi, R. Y.; Soboh, B.; Curatti, L.; Ludden, P. W.; Dean, D. R.; Rubio, L. M. *Mol. Microbiol.* **2007**, *63*, 177–192.
- (17) We note that the nonstoichiometric quantitation of Mo in our NifEN samples allows the possibility that Mo bound to the NifB-co derivative is simply not extracted by NifX. However, as NifX readily binds FeMo-co as well as NifB-co, this possibility seems unlikely.
- (18) Beinert, H.; Thomson, A. J. *Arch. Biochem. Biophys.* **1983**, *222*, 333–361.
- (19) For example, see: Butt, J. N.; Armstrong, F. A.; Breton, J.; George, S. J.; Thomson, A. J.; Hatchikian, E. C. *J. Am. Chem. Soc.* **1991**, *113*, 6663–6670.

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